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## Review on role of nrf2 pathway activation in neurological disorder

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Article History	Abstract
Received on: 15-11-2022 Revised on: 22-11-2022 Accepted on: 27-12-2022	With protein build-up and mitochondrial damage leading to brain problems, oxidative stress is a key factor in the onset of many neurodegenerative illnesses. Several defence systems, including nuclear erythroid factor2 (Nrf2)-Kelch-like ECH-associated protein1, protect nerve cells by producing antioxidants to reduce oxidative stress (Keap1) In a number of neurological illnesses, signalling pathway activation has been shown to be a promising therapeutic for reducing oxidative stress and neuroinflammation and protecting neurons. In this review, we specifically highlight Nrf2's beneficial effects on Alzheimer's and Parkinson's disorders. By releasing over 250 cytoprotective genes intended to combat oxidative stress and neuroinflammation, Nrf2 has demonstrated that it is a master regulator of antioxidants. It has been demonstrated in animal experiments that Nrf2 activation enhances autophagy, mitochondrial biogenesis, and the suppression of inflammatory cytokinin which protects neuronal cells and inhibit progressive neurodegeneration.
<b>Keywords:</b> Oxidative stress, Neurodegeneration, Nrf2, Parkinson's disease, Antioxidant, Keap1	
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### Introduction

The human brain consists of about 1011–1012 neuronal cells which is maintained by neuroglial cells such as oligodendrocytes, astrocytes, and microglia. Recent research proven a strong linkage between inflammation and neuronal cell death; moreover, this would be a reason this scenario is unclear that it would be a direct cause of neurodegeneration [1]. Despite neurological disorders linked with common risk factors, such as oxidative stress, misfolded proteins, aging, and

environmental hazards, this exhibits microglial activation and mitochondrial dysfunction in each disease progression by damage to lipids, proteins, and DNA [2]. Misfolded and aggregation proteins are a similar feature in many neuronal disorders even though misfolded proteins are inactive, but stress and endoplasmic reticulum (ER) stress. Unfolded protein response (UPR) generally maintained by ER gets altered and promotes protein misfolding and reduces ER protein levels by proteasomal degradation and autophagy [3]. In the central nervous system (CNS), there is a very limited rate of regeneration for neurons and they are sensitive to oxidative stress so neurodegeneration must be controlled in the brain. Neuronal loss seen mostly by apoptosis or necrosis. In

apoptosis, an intrinsic suicide program initiates by cascade for cellular death. In necrosis accidental cell death with uncontrolled release of inflammatory mediators [4]. The most common neurodegenerative disorders are Alzheimer's disease (AD), which shows marked neuronal loss by an accumulation of misfolded amyloid plaques, which increases with aging in the hippocampus and the frontal cortex region and secondary to chronic inflammation that result from amyloid accumulation links directly with oxidative stress ultimately leads to severe memory loss [5]. In Parkinson's disease (PD), inflammation results in the loss of dopaminergic neurons in substantia nigra resulting in an unusual hypokinetic movement disorder. However, activation of inflammatory cytokines is essential for the self-defense mechanism of CNS toward foreign antigens but prolonged expression of inflammatory mediators may lead to cell death [6]. The purpose of this review is to focus on two major neurological problems of AD and PD and the neuroprotective effect of nuclear erythroid factor2 (Nrf2) against oxidative stress has been significantly proven in many neurodegenerative animal models. Nrf2 enhances many transcriptional genes of cytoprotective and Phases I and II drug detoxifying enzymes, and mitochondrial pathways in addition to affecting various antioxidant enzymes can also boost the expression of anti-inflammatory mediators, which shows neuroprotective activity [7]. Cortical culture has shown that Nrf2 knockout mice are vulnerable to oxidative stress, and Nrf2 expression enhances neuroprotection in PD and AD through antioxidative response elements (ARE) activation [8].

**Nrf2 Structure**

Nrf2 was discovered in 1994 at the laboratory of YuetWaiKan and it belongs to the cap "n" collar (CNC) subfamily of basic leucine zipper (bZIP) transcription factors and is encoded by the NFE2L2 gene [9,10].

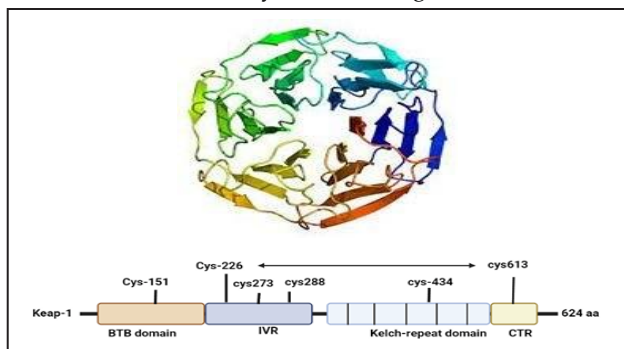


Figure 1. Human Nrf2 Protein Structure.

It includes members with a CNC homology region of 43 amino acids, which links the N-term DNA-binding domain to the family's DNA-binding specificity [11]. Nrf2 is a complex protein that contains seven functional domains. Neh 1-7 each of which has a specific function, as shown in Figure 1.10,11 Plafker et al [16]. discovered that Neh1 has a bZIP that forms a dimer from two different monomers with tiny musculoaponeurotic fibrosarcoma protein (maf), DNA, and other transcription partners. It has been discovered that Nrf2's Neh1 forms a nuclear complex containing the ubiquitin-conjugating enzyme UbcM2, which controls Nrf2 stability through DNA bindin [12, 13]. Neh2 N-terminus interacts with the kelch domain of Keap1 (Cullin (Cul)3-RING), dimeric redox-sensitive substrate adaptor Ubiquitin ligase complex including the box protein (Rbx)1 (i.e., CRLKeap1) via two binding sites, the more powerful ETGE region and the weaker DLG region [9,14,15] shown in Figure 2. Interaction of the C-terminal Neh3 with the transcription coactivator CHD6 a chromo-ATPase/helicase DNA-binding protein may play a critical role in the activation of ARE-driven genes, according to research [16]. Neh4 and Neh5 bind together to CREB-binding protein (CBP), synergistically stimulate gene expression via ARE, and synchronize reporter gene activation [17]. Neh6 includes two motifs: (1) DSGIS and (2) DSAPGS binding to dimericbTrCP (b-transducin repeat-containing protein), and S-phase kinase- associated protein acts as the main receptor for this protein, 1 (Skp1)-Cul1-Rbx1 core E3 complex depicted in Figure 2. While Neh6 controls Nrf2 stability without engaging with Keap1, Neh7 inhibits the Nrf2-ARE signaling pathway by connecting with the retinoid X receptor alpha (RXR) [14-18].

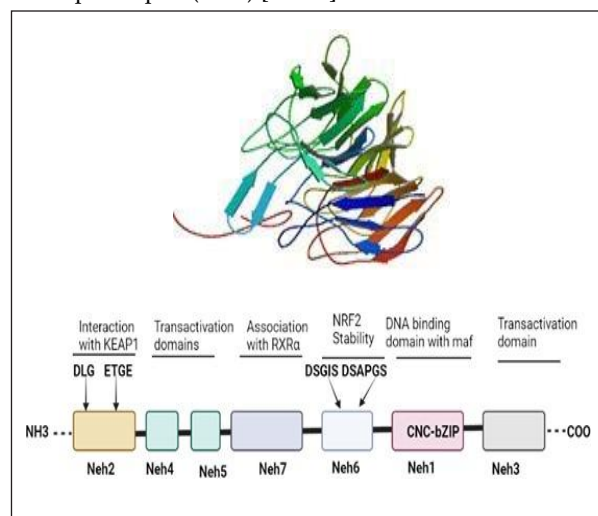


Figure 2. KEAP-1 Protein.

### Keap1–Nrf2–ARE Pathway

The Keap1–Nrf2 pathway is a crucial antioxidant signaling pathway against reactive oxygen species, which is one of the major critical players in chronic inflammation associated with PD and AD. The antioxidant enzyme genes glutathione S-transferase (GST) and NAD(P)H quinone oxidoreductase1 (NQO1), as well as heme oxygenase-1 (HO-1), are all enhanced by Nrf2 and induction of an ARE DNA sequence [18,19]. Keap1 suppresses the activity of Nrf2 by binding with Neh2, Keap1 act as an adapter molecule for the Cul3 E3 ligase complex, causing Nrf2 to be degraded via the ubiquitin-proteasome pathway, exposure to electrophiles modifies or reactive oxygen species Keap1 inactivates thereby leading to activation of Nrf2 [20]. Causing accumulation of Nrf2 in the nucleus and heterodimerization with small Maf proteins binds to ARE at the regulatory region of the targeted gene [21, 22] shown as in figure 3.

In unstressed conditions, Nrf2 protein level is the low reason being Keap1/Cul3 E3 ubiquitin ligase complex degrades proteasomal proteins [23, 24]. Reactive oxygen species modify specific cysteine residue of Keap1 (C257, C273, C288, and C297) causing disassociation of Keap1 and Nrf2 migrate to nucleus causing its accumulation as evidenced in Keap1 knockdown in human cells and KEAP1 deletion in mice, and activation of cytoprotective genes [23–26]. Many cytoplasmic proteins that have Keap1 dependent activity that stabilizes Nrf2 by attenuating interaction between Keap1- Nrf2. P62/sequestosome 1 (SQSTM1) also known as Autophagy cargo-adaptor, [27–32] dipeptidyl peptidase 3 (DPP3), [31] Wilms tumor gene on the X chromosome (WTX), [33] and Localizer of BRCA2 (PALB2) [34] because they all possess Keap1-interacting region (KIR)-like ETGE motifs, they interact with Nrf2 for Keap1 attachment, allowing Keap1 to be sequestered and Nrf2 to be maintained EDGE suppresses DLG via interacting with the P21 cyclin-dependent kinase p21Cip1/WAF1 [35].

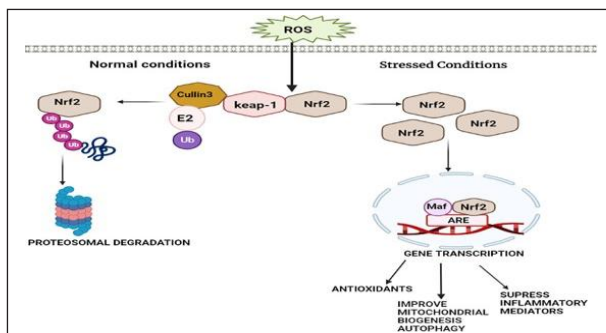


Figure 3. The Mechanism of Nrf2 Activation.

BRCA1 promotes stability and activation of Nrf2 P300 interacts with Nrf2 and interferes with Keap1- Nrf2 complex formation [37] which leads to Nrf2 activation and enhance four genes, including Malic enzyme1, Glucose 6-phosphate dehydrogenase, Isocitrate dehydrogenase, and 1,6-phosphogluconate dehydrogenase all of which play a role in the production of NADPH, an antioxidant cofactor and fuel. It moreover regulates cytochrome P450 oxidoreductase activity. Neutrophils and monocytes have the highest levels of NRF2 among blood cells. Nrf2 levels are high in astrocytes. Furthermore, microglia, a kind of monocyte, have larger levels of Nrf2 than neurons [37–40]. Nrf2 reduces inflammation in three ways modifying redox metabolism, interacting with (NF-KB), and directly regulating pro-inflammatory mediators responsible for neurodegeneration in PD and AD [40,41].

### Nrf2 and Mitochondria

Mitochondria consist of membranes which are separating them from other cellular organelles was thought to be responsible only for energy generation due to their genetic autonomy, however, studies suggest that a transduction network in the cytosol circuit of mitochondria maintains cellular equilibrium [42]. Nrf2 controls mitochondrial biogenesis, and autophagy both directly or indirectly [43] Nrf1 controls mitochondrial transcription factors when it is linked to peroxisome proliferator-activated receptor-gamma coactivator 1-alpha (PGC1) [44] is a gene that controls mitochondrial development. To maintain mitochondrial integrity and function, continuous monitoring and controlled autophagosome clearance of damaged mitochondria are necessary. Under oxidative stress, Nrf2 increases PTEN-induced putative kinase protein 1 (PINK1) expression, which controls mitochondrial efficiency [45] Mitosis in mitochondria serves to preserve PINK1 on the outer membrane of damaged mitochondria, which Parkin subsequently uses to activate P62, causing mitochondrial aggregation and eventual elimination [46]. Furthermore, it has been discovered that increasing mitophagy enhances mitochondrial excitation and minimizes a buildup of protein aggregates induced neurotoxicity. By boosting Nrf2-mediated mitophagy and blocking the mTOR pathway, sulforaphane rescued animals with experimental PD from rotenone-induced DA neurodegeneration [47]. In neurodegenerative diseases, Nrf2 activation has a beneficial influence on mitochondrial dysfunction has been supported by a large body of experimental evidence. MitoQ is a

traumatic brain damage mouse model, a mitochondrial-targeted antioxidant corrected neurological impairments. Increases in mitochondrial activity and antioxidant gene expressions, such as superoxide dismutase enzyme (SOD1) and catalase, are mediated by the activation of Nrf2 by free radicals. The Glutathione-S-transferases (GSHs) system is mediated by the Nrf2/ARE pathway which is important for maintaining mitochondrial-generated reactive oxygen species (ROS) and is altered in PD and AD pathogenesis. Cell lines generated from PD patients, which contain reduced GSH levels, can also be restored by activating Nrf2. Overall, there is significant evidence that targeting the Nrf2/ARE pathway in neurodegenerative illnesses such as in PD and AD enhances mitochondrial function and lowers excessive mitochondrial oxidative stress [48].

#### **Role of Nrf2 in Mitochondrial Dysfunctions**

The aging mechanism is linked with mitochondrial failure and ROS. The discovery of neurotoxic compound 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) is an accidental human exposure to induced mitochondrial failure was the first proof that mitochondrial dysfunction plays a role in PD, further studies are needed to understand the mechanism of MPTP induced dopaminergic neurotoxicity by converting into 1-methyl-4-phenylpyridinium (MPP+) in brain, damages complex I in mitochondria resulting in mitochondrial ROS and oxidative stress leads to release of cytochrome c reductase in the mitochondrial respiratory chain further lead to mitochondrial failure [49]. Low concentrations of complex I subunit and its activity have been found in postoperative PD individual midbrain biopsies and patient-derived platelets.<sup>50</sup> As a result, the experimental shreds of evidence show that aging reduces cellular protective properties by weakening the Nrf2/ARE pathway's role. Nrf2-Keap1 protein complex functions as a cellular electrochemical detector, regulating antioxidant gene transcription to maintain redox homeostasis. The Nrf2/ARE activity is influenced by both aging and neurodegenerative disorders. In multiple tissues, a meta-analysis of PD and AD identified 54 impacted genes, 31 of which contain ARE. GSR1 promotes glutathione reduction by employing the reducing power of nicotinamide adenine dinucleotide diphosphate as a subunit of glutamate-cysteine ligase (NADPH).<sup>49,51</sup> Nrf2 regulates the transcription of aldehyde dehydrogenases, which are essential in alcohol elimination and reduce NADP+ to NADPH. DMF and its bioactive metabolite, monomethylfumarate (MMF), have been demonstrated to accelerate mitochondrial

oxidative phosphorylation and biogenesis by activating Nrf2 [52]. As a result, the Nrf2/ARE system influences both mitochondrial and non-mitochondrial metabolic pathways, indicating therapeutic potential in neurodegenerative diseases.

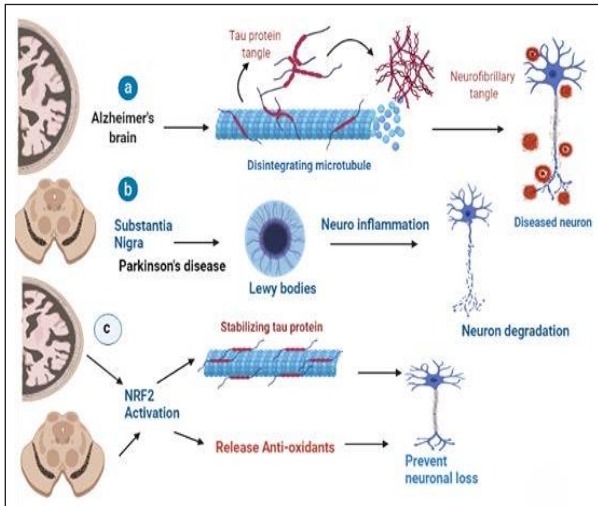
#### **Nrf2 Expression in Neurodegenerative Diseases**

Unlike many tissues, Nrf2 is released at elevated levels in the liver, kidneys, and lungs. Moreover, it is released at a greater level in gastrointestinal tissues (esophagus, stomach, small intestine, and large intestine), studies conducted by Chan et al.<sup>9</sup> reported that the expression of Nrf2 is seen in the lateral ventricle and its mid-wall, as well as in the choroid plexus of the fourth ventricle in the brain, Nrf2 is also observed in the nasal cavity tissue involved in smell, thyroid, submandibular glands, and the brown fat [5].

In AD, Nrf2 is primarily expressed in cytoplasmic hippocampus neurons; it does not translocate to the nucleus; a biochemical extract of the frontal cortex confirmed decreased levels of nuclear Nrf2 in AD. In the nuclei of AD patients, neither neurons nor astrocytes showed substantial Nrf2 staining. The Nrf2 pathway is most likely dysfunctional in AD hippocampal neurons; nevertheless, in PD, Nrf2 is more numerous in dopamine neurons than in SNpC, and dopaminergic Nrf2 exhibits neuronal sensitivity to increased oxidative stress [6].

#### **Role of Nrf2/ARE Pathway in AD**

In the last few decades, many studies have discovered different features of Nrf2 expression and relevant antioxidant genes in AD models, a considerable increase in nuclear Nrf2 levels was seen in investigations. The existence of cytoplasmic confinement in the AD patient's hippocampus was discovered as indicating a reduction in levels of the Nrf2 gene [53]. In AD hippocampal brain areas, NQO1 activity was upregulated. Similarly, the activities of catalase and SOD1 in the frontal lobe and temporal cortex of the AD brain appeared to be diminished.<sup>54</sup> Mishaps in the Nrf2 pathway are associated with peripheral tissues as a reflection of process going on in the brain and spinal cord. Reduced cognitive impairments, oxidative damage, and inflammatory processes in AD models, and counteract against a mediated inflammation-induced, multiple mechanisms involved in the activation of Nrf2 have been identified in AD. The possibility of targeting Nrf2-regulatory proteins in the therapy of AD is demonstrated in this section [55].



**Figure 4.** The Neuroprotective Role of Nrf2 Activation in Both AD and PD Neurodegradation.

p62, a polyubiquitination protein, can promote Nrf2 activity by identifying Keap1 as a target for degradation. Different researchers have discovered that there is a link among Keap1, p62, and Nrf2, which indicates a reduction of protein p62 results decreased Keap1 breakdown and as a consequence, increase in the release of Nrf2, therefore, A $\beta$  insertion into the rat's hippocampal region enhanced catabolic proteins including LC3-II and beclin1, as well as Keap1, while reduced p62 and Nrf2 in the cortex and hippocampus.<sup>56</sup> In the cortex region of Alzheimer's brains, however, mRNA and gene expression of p62, and also Nrf2 target genes, were found to be elevated [57]. Several investigations demonstrated that Nrf2 activity is decreased in 3xTg-AD mouse model begins to develop an age-dependent and transformative neurodegenerative genetic makeup, which includes an age-dependent plaque and tangles in the brain, similar to AD patients, due to three protein abnormalities in presenilin-1, amyloid precursor protein, and microtubule-linked tau which never interact in human AD kinship forms [58]. From 3 to 5 months of age, the 3xTg-AD mice reported lower Glutathione and vitamin E levels, but also enhanced GSH-Px activities. Furthermore, 3-month-old PBMC from 3xTg-AD exhibited elevated p (Ser40)Nrf2 which corresponded to Nrf2 genetic translation Sensor same as human MCI PBMCs.<sup>59</sup> In the AD animal model PS1V97LTg, with a change in the PSEN1 gene, the Nrf2 inducer sulforaphane promoted enhanced Nrf2 activation in the brain and the Nrf2-ARE pathway alleviated oxidative imbalance amyloidopathy and improves cognitive performance. When compared to mice of the mutant strain studies shows amyloid proteins build and

aggregation of tau protein into neurofibrillary with Nrf2-leakage (AT-Nrf2-KO) that shows an abnormal increase in the number of astrocytes and accumulation of microglial cells as a reaction to injury (AT-Nrf2-WT) see Figure 4 [60].

#### Role of Nrf2/ARE Pathway in PD

Ramsey et al.<sup>10</sup> investigated higher concentrations of Nrf2 in the nucleus of human PD substantial nigra neurons. Multiple investigations have correlated the Nrf2 pathway to various PD models. Interestingly, results from individual samples show that Nrf2 nuclear translocation significantly improved [61–65]. Suggesting a higher level of transcriptional regulation than it was in age-matched subjects GSH concentrations were also observed to be decreased in the substantia nigra of individuals with PD brains, whereas HO-1, NQO1 levels were shown to be elevated in astrocytes, endothelial cells, and dopamine neurons from PD autopsy brains [66]. In PD brains, forensic researches reveal that Nrf2 nuclear translocation is significantly greater, along with a higher concentration in certain Nrf2-regulated proteins.  $\alpha$ -Syn was stereotactically delivered in the ventral midbrain of Nrf2/mice using adeno-associated viral vectors to properly understand the purpose of Nrf2 in PD.<sup>67</sup> In Nrf2/mice treated with MPTP or 6-OHDA, which promote oxidative stress and are frequently used to imitate PD in rats, identical results were obtained. Additionally, transplantation astrocytes Nrf2 overexpression into the striatum of wild-type mice reduced sensitivity to 6-OHDA and MPTP during 6-OHDA treatment, and parallel values were demonstrated in cortical neurons from Nrf2+/- animals uncontrolled release of Nrf2 in astrocytes with mice uncontrolled expressing human mutation SYN (hSYNA53T) in neurons resulted in delayed motor dysfunction, lower Synuclin protein buildup, and attenuate ROS along with diminished microglia in the spinal cord see Figure 4.<sup>68</sup> Mild regular physical activity was demonstrated beneficial in mice treated with 6-OHDA through Nrf2 neuron protective action, boosting mitochondrial biogenesis acceleration and limiting Parkinsonism progress [69]. As a result, impaired mitochondrial mitophagy, efficiency, and biogenesis are major pathogenic characteristics of PD, and Nrf2 is a signaling pathway factor that controls mitochondrial biogenesis checks and homeostasis. As a result, there are many shreds of evidence of the role of Nrf2 in the pathogenesis of PD, indicating that increasing the Nrf2/ARE pathway as an effective PD treatment is of interest [70]. As a result, increased p62 availability,

which might potentially lead to increased Nrf2 target gene expression, could be a promising therapy for PD. Despite this, there have been no investigations on the usage of p62 oscillators and their influence on Nrf2 transmission in PD. Glycogen synthase kinase 3 (GSK-3) kinase will suppress Nrf2, leading to reduced Nrf2 activity due to increased degradation. Furthermore, there is evidence that GSK-3 activation is modified in PD. Intracellular GSK-3 levels were upregulated with Syn in Lewy bodies from PD human brains in the brainstem and upper parietal lobe, as well as the striatal and inferior frontal lobe [71]. GSK-3 inhibition, PI3K/Akt stimulation, and elevated Nrf2 expression of target genes, PC12 cells, and SH-SY5Y cells reduced oxidative imbalance caused by the cytotoxic agents of MPTP, MPP+, and inhibited apoptosis. Berberine an isoquinoline compound exhibiting PI3K-activating action, also attenuated 6-OHDA-induced dopaminergic neuron loss and motor movement disturbances in zebrafish via enhancing PI3K/Akt activation and Nrf2 activation [72]. Overall, in multiple PD models, manipulation of the PI3K/Akt/GSK-3 pathway and restriction of the p38 MAPK pathway seemed to be the most viable target pathways, leading to Nrf2 activation and subsequent stress response. Minimizing neurotoxicity and promoting the Nrf2/ARE pathway should be an area to explore in PD treatment, based on studies in PD postmortem tissues [73].

### Conclusion

ROS or superoxides react with lipids, proteins, and DNA resulting in neuron cell death, which is the primary cause of neurological diseases including PD and AD. The parameters like protein buildup, mitochondrial dysfunction, and proteasomes dysfunction is also reasons that may cause cell death and neurological diseases. Nrf2 pathway activation can help to reduce oxidative stress and maintain mitochondrial biogenesis while also increasing autophagy, which helps to destroy misfolded proteins. In this review, it was focused on the importance of Nrf2 pathway activation that has been proved with scientific studies in neurological diseases. The study suggests that the PI3K/Akt/GSK-3 pathway, which looks to be well-studied for AD and PD therapies, is involved in the positive regulation of Nrf2 activity. Restriction of the NF- $\kappa$ B pathway must also be investigated as a prospective AD and PD therapeutic goal. The evidence clearly states that drugs that activate Nrf2, including

those that have been proven to elevate Nrf2-related genes, have a protective effect on AD and PD models.

### References

1. Emerit J, Edeas, M, and Bricaire, F. Neurodegenerative diseases and oxidative stress. *Biomed Pharmacother* 2004; 58(1): 39–46.
2. Coppedè F and Migliore, L. DNA damage in neurodegenerative diseases. *Mut Res/Fundament Mol Mech Mutag* 2015; 776: 84–97.
3. Hetz C. The unfolded protein response: controlling cell fate decisions under ER stress and beyond. *Nat Rev Mol Cell Biol* 2012; 13(2): 89–102.
4. Kanduc D, Mittelman, A, and Serpico, R, et al. Cell death: Apoptosis versus necrosis (Review). *Int J Oncol* 2002; 21(1):165–70.
5. Cole, GM, Lim, GP, and Yang, F, et al. Prevention of Alzheimer's disease: Omega-3 fatty acid and phenolic antioxidant interventions. *Neurobiol Aging* 2005; 26(1): 133–136.
6. Campbell, A. Inflammation, neurodegenerative diseases, and environmental exposures. *Ann NY Acad Sci* 2004; 1035(1): 117–132.
7. Uppugalla S, Male U, Srinivasan P. Design and synthesis of heteroatoms doped carbon/polyaniline hybrid material for high performance electrode in supercapacitor application. *ElectrochimicaActa*. 2014 Nov 10;146:242-8.
8. Kraft, AD. Nuclear factor E2-related factor 2-dependent anti-oxidant response element activation by tert-butylhydroquinone and sulforaphane occurring preferentially in astrocytes conditions neurons against oxidative insult. *J Neurosci* 2004; 24(5): 1101–1112.
9. Chan K, Lu, R, and Chang, JC, et al. NRF2, a member of the NFE2 family of transcription factors, is not essential for murine erythropoiesis, growth, and development. *Proc Natl Acad Sci* 1996; 93(24): 13943–13948.
10. Male U, Uppugalla S, Srinivasan P. Effect of reduced graphene oxide-silica composite in polyaniline: electrode

- material for high-performance supercapacitor. *Journal of Solid State Electrochemistry*. 2015 Nov;19(11):3381-8.
11. Moi P, Chan, K, and Asunis, I, et al. Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the beta-globin locus control region. *Proc Natl Acad Sci* 1994; 91(21): 9926–9930.
  12. Cuadrado A, Rojo AI, and Wells, G, et al. Therapeutic targeting of the NRF2 and KEAP1 partnership in chronic diseases. *Nat Rev Drug Disc* 2019; 18(4): 295–317. <https://www.nature.com/articles/s41573-018-0008-x>
  13. Song M-Y, Lee D-Y, Chun, K-S, et al. The role of NRF2/ KEAP1 signaling pathway in cancer metabolism. *Int J Mol Sci* 2021; 22(9): 4376.
  14. Singh, A., 2022. Hyperlipidemia in cardiovascular health and digestion. In *Nutrition and Functional Foods in Boosting Digestion, Metabolism and Immune Health* (pp. 141-150). Academic Press.
  15. Namani A, Li Y, and Wang, XJ, et al. Modulation of NRF2 signaling pathway by nuclear receptors: implications for cancer. *Biochimica et Biophysica Acta (BBA) – Mol Cell Res* 2014; 1843(9): 1875–1885.
  16. Plafker, KS, Nguyen, L, and Barneche, M, et al. The ubiquitin-conjugating enzyme UbcM2 can regulate the stability and activity of the antioxidant transcription factor Nrf2. *J Biol Chem* 2010; 285(30): 23064–23074.
  17. Tong KI, Katoh Y, and Kusunoki, H, et al. Keap1 recruits Neh2 through binding to ETGE and DLG motifs: characterization of the two-site molecular recognition model. *Mol Cell Biol* 2006; 26(8): 2887–900.
  18. Hayes JD and Dinkova-Kostova AT. The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem Sci* 2014; 9(4): 199–218. <https://pubmed.ncbi.nlm.nih.gov/24647116/>
  19. Nioi P, Nguyen T, and Sherratt PJ, et al. The carboxy-terminal Neh3 domain of Nrf2 is required for transcriptional activation. *Mol Cell Biol* 2005; 25(24): 10895–10906. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1316965>
  20. Uppugalla S, Srinivasan P. High-performance supercapacitor coin cell: polyaniline and nitrogen, sulfur-doped activated carbon electrodes in aqueous electrolyte. *Journal of Solid State Electrochemistry*. 2019 Jan;23(1):295-306.
  21. Kim S, Indu Viswanath AN, Park, J-H, et al. Nrf2 activator via interference of Nrf2-Keap1 interaction has antioxidant and anti-inflammatory properties in Parkinson's disease animal model. *Neuropharmacology* 2020; 167: 107989.
  22. Ma Q. Role of Nrf2 in oxidative stress and toxicity. *Ann Rev Pharmacol Toxicol* 2013; 53: 401–426. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4680839/>
  23. Uppugalla S, Srinivasan P. Polyaniline nanofibers and porous Ni [OH] 2 sheets coated carbon fabric for high performance super capacitor. *Journal of Applied Polymer Science*. 2019 Nov 5;136(41):48042.
  24. Stępkowski TM and Kruszewski MK. Molecular cross-talk between the NRF2/KEAP1 signaling pathway, autophagy, and apoptosis. *Free Radic Biol Med* 2011;50(9):1186–95. doi: 10.1016/j.freeradbiomed.2011.01.033
  25. Singh, A., 2022. Role of microbial metabolites in cardiovascular and human health. In *Microbiome, Immunity, Digestive Health and Nutrition* (pp. 137-148). Academic Press.
  26. Roh, J., Hill, J.A., Singh, A., Valero-Muñoz, M. and Sam, F., 2022. Heart failure with preserved ejection fraction: heterogeneous syndrome, diverse preclinical models. *Circulation Research*, 130(12), pp.1906-1925.
  27. Baird, L, Llères, D, and Swift, S, et al. Regulatory flexibility in the Nrf2-mediated stress response is conferred by conformational cycling of the Keap1-Nrf2 protein complex. *Proc Natl Acad Sci* 2013; 110(38): 15259–15264. <https://www.pnas.org/content/110/38/15259.short>

28. Wakabayashi, N, Itoh, K, and Wakabayashi, J, et al. Keap1-null mutation leads to postnatal lethality due to constitutive Nrf2 activation. *Nat Genet* 2003; 5(3): 238–245. <https://www.nature.com/articles/ng1248/briefing/signup/?origin=Nature&originReferralPoint=EmailBanner>
29. Devling, TWP, Lindsay, CD, and McLellan, LI, et al. Utility of siRNA against Keap1 as a strategy to stimulate a chemopreventive phenotype. *Proc Natl Acad Sci* 2005; 102(20): 7280–7285.
30. Taniguchi, K, Yamachika, S, and He, F, et al. p62/SQSTM1-Dr. Jekyll and Mr. Hyde that prevents oxidative stress but promotes liver cancer. *FEBS Lett* 2016; 590(15): 2375–2397.
31. Uppugalla S, Boddula R, Srinivasan P. Methyl triphenylphosphonium permanganate as a novel oxidant for aniline to polyaniline-manganese (II, IV) oxide: material for high performance pseudocapacitor. *Journal of Solid State Electrochemistry*. 2018 Feb;22(2):407-15.
32. Umemura, A, He, F, and Taniguchi, K, et al. p62, Upregulated during preneoplasia, induces hepatocellular carcinogenesis by maintaining survival of stressed HCC-initiating cells. *Cancer Cell* 2016; 29(6): 935–948.
33. Hast, BE, Goldfarb, D, and Mulvaney, KM, et al. Proteomic analysis of ubiquitin ligase KEAP1 reveals associated proteins that inhibit NRF2 ubiquitination. *Canc Res* 2013; 73(7): 2199– 2210.
34. Parajuli D, Uppugalla S, Murali N, Ramakrishna A, Suryanarayana B, Samatha K. Synthesis and Characterization MXene-Ferrite Nanocomposites and its application for Dying and Shielding. *Inorganic Chemistry Communications*. 2022 Dec 16:110319.
35. Lau, A, Wang, X-J, and Zhao, F, et al. A noncanonical mechanism of Nrf2 activation by autophagy deficiency: direct interaction between Keap1 and p62. *Mol Cell Biol* 2010; 30(13): 3275–3285.
36. Camp, ND, James, RG, and Dawson, DW, et al. Wilms tumor gene on X chromosome (WTX) inhibits degradation of NRF2 protein through competitive binding to KEAP1 protein. *J Biol Chem* 2012; 287(9): 6539–6550.
37. Singh, A., Kumar, A. and Kalaiselvi, P., 2018. Aegeline, targets LOX1, the receptor for oxidized LDL to mitigate hypercholesterolemia: a new perspective in its anti-atherosclerotic action. *Free Radical Biology and Medicine*, 128, p.S41.
38. Botsa SM, Seetharam P, Raju IM, Suresh P, Satyanarayana G, Sambasivam S, Susmitha U, Tejeswararao D. Nanohybrid material of Co–TiO2 and optical performance on methylene blue dye under visible light illumination. *Hybrid Advances*. 2022 Dec 8:100008.
39. Gorrini, C, Baniasadi, PS, and Harris, IS, et al. BRCA1 interacts with Nrf2 to regulate antioxidant signaling and cell survival. *J Exp Med* 2013; 210(8): 1529–1544.
40. Ganner, A, Pfeiffer, Z-C, and Wingendorf, L, et al. The acetyltransferase p300 regulates NRF2 stability and localization. *Biochem Biophys Res Commun* 2020; 524(4): 895–902.
41. Akmal A, Javaid A, and Hussain R, et al. Screening of phytochemicals against Keap1- NRF2 interaction to reactivate NRF2 Functioning: Pharmacoinformatics based approach. *Pak J Pharm Sci* 2019 ;32(6(Supplementary)):2823–2828.
42. Lee, J-M, Calkins, MJ, and Chan, K, et al. Identification of the NF-E2-related factor-2-dependent genes conferring protection against oxidative stress in primary cortical astrocytes using oligonucleotide microarray analysis. *J Biol Chem* 2003; 278(14): 12029–12038.
43. Uppugalla S, Rajesh K, Surendra AV, Kumar K, Gayasuddin M. Effect Of Pisonia Alba Root Extract On Cafeteria Diet-Induced Obesity In Rats. *Journal of Pharmaceutical Negative Results*. 2022 Dec 1:3732-9.
44. Mitsuishi, Y, Taguchi, K, and Kawatani, Y, et al. Nrf2 redirects glucose and glutamine into anabolic pathways in metabolic reprogramming. *Canc Cell* 2012; 22(1): 66–79.

45. Barbeau, A, Dallaire, L, and Buu, NT, et al. Comparative behavioral, biochemical and pigmentary effects of MPTP, MPP+ and paraquat in rana pipiens. *Life Sci* 1985; 7(16): 1529–1538.
46. Yun, J and Finkel, T. Mitohormesis. *Cell Metab* 2014; 19(5): 757–766.
47. Hayashi, Y, Regnier, T, and Nishiguchi, S, et al. ChemInform abstract: Efficient total synthesis of (+)-Negamycin, a potential chemotherapeutic agent for genetic diseases. *ChemInform* 2008; 9(38): 2379–81. doi: 10.1039/b801498a
48. Mahajan, A, and Ahuja, A. Authors' reply to Tandon et al., Kudva et al., and Krishnan et al. *Canc Res Stat Treat* 2020; (2): 412.
49. Sriram N, Uppugalla S, Rajesh K. Cognitive Enhancing And Antioxidant Activity Of Ethyl Acetate Soluble Fraction Of The Methanol Extract Of Pisonia Alba Leaves In Scopolamine-Induced Amnesia. *Journal of Pharmaceutical Negative Results*. 2022 Dec 1:3740-9.
50. Monach, PA. Global versus organ-specific outcome measures in systemic lupus erythematosus: comment on the articles by Furie et al, Nikpour et al, Wallace et al, Burgos et al, and Ramos-Casals et al. *Arthrit Care Res* 2010; 62(4): 580–581.
51. Wang S, Wang Z, and Gao, H, et al. Highly regioselective palladium-catalyzed domino reaction for post-functionalization of BODIPY. *Chem Commun (Camb)* 2021;57(14):1758–1761. doi: 10.1039/d0cc08163a
52. Singh, A., Srinivasan, A.K., Chakrapani, L.N. and Kalaiselvi, P., 2019. LOX-1, the common therapeutic target in hypercholesterolemia: a new perspective of antiatherosclerotic action of aegeline. *Oxidative medicine and cellular longevity*, 2019.
53. Ammal Kaidery, N, Ahuja, M and Thomas, B. Crosstalk between Nrf2 signaling and mitochondrial function in Parkinson's disease. *Mol Cell Neurosci* 2019; 101: 103413.
54. Bing, RG, Sulis, DB, and Wang, JP, et al. Thermophilic microbial deconstruction and conversion of natural and transgenic lignocellulose. *Environ Microbiol Rep* 2021; 13(3): 272–293.
55. Osama A, Zhang J, and Yao, J, et al. Nrf2: A dark horse in Alzheimer's disease treatment. *Ageing Res Rev* 2020;64:101206. doi: 10.1016/j.arr.2020.101206
56. Shiv Chandra Singh, A., Yu, A., Chang, B., Li, H., Rosenzweig, A. and Roh, J.D., 2021. Exercise Training Attenuates Activin Type II Receptor Signaling in the Aged Heart. *Circulation*, 144(Suppl\_1), pp.A14259-A14259.
57. Dickenson, et al. v. Ramsey et al. Nov. 20, 1913 [79 S. E. 1025]. *The Virginia Law Register* 1914; 19(11): 845.
58. Conrad, B. In Response to Murphy et al., Löwer et al., and Lan et al. *Cell* 1998; 95(1): 16.
59. Zhang Z, Li G, and Szeto, SSW, et al. Examining the neuroprotective effects of protocatechuic acid and chrysin on in vitro and in vivo models of Parkinson disease. *Free Rad Biol Med* 2015; 84: 331–343.
60. Tanji K, Odagiri S, and Miki, Y, et al. p62 Deficiency enhances  $\alpha$ -synuclein pathology in mice. *Brain Pathol* 2014; 25(5): 552–564.
61. Oddo, S, Caccamo, A, and Shepherd, JD, et al. Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular A $\beta$  and synaptic dysfunction. *Neuron* 2003; 9(3): 409–421. <https://www.ncbi.nlm.nih.gov/pubmed/12895417>
62. Singh, A., Gowtham, S., Chakrapani, L.N., Ashokkumar, S., Kumar, S.K., Prema, V., Bhavani, R.D., Mohan, T. and Sathyamoorthy, Y.K., 2018. Aegeline vs Statin in the treatment of Hypercholesterolemia: A comprehensive study in rat model of liver steatosis. *Functional Foods in Health and Disease*, 8(1), pp.1-16.
63. Tian, Y, Wang, W, and Xu, L, et al. Activation of Nrf2/ARE pathway alleviates the cognitive deficits in PS1V97L-Tg mouse model of Alzheimer's disease through modulation of oxidative stress. *J Neurosci Res* 2018; 97(4): 492–505.

64. Rojo, AI, Pajares, M, and García-Yagüe, AJ, et al. Deficiency in the transcription factor NRF2 worsens inflammatory parameters in a mouse model with combined tauopathy and amyloidopathy. *Redox Biol* 2018; 18: 173–180.
65. Zgorzynska E, Dziedzic B, and Walczewska A. An overview of the Nrf2/ARE pathway and its role in neurodegenerative diseases. *Int J Mol Sci* 2021;22(17):9592. doi: 10.3390/ijms22179592
66. van Muiswinkel, FL, de Vos, RAI, and Bol, JGJM, et al. Expression of NAD(P)H:quinone oxidoreductase in the normal and Parkinsonian substantia nigra. *Neurobiol Aging* 2004; 25(9): 1253–1262.
67. Lastres-Becker, I, Ulusoy, A, and Innamorato, NG, et al.  $\alpha$ -Synuclein expression and Nrf2 deficiency cooperate to aggravate protein aggregation, neuronal death and inflammation in early-stage Parkinson's disease. *Human Mol Genet* 2012; 21(14): 3173–3192.
68. Gan, L, Vargas, MR, and Johnson, DA, et al. Astrocyte-specific overexpression of Nrf2 delays motor pathology and synuclein aggregation throughout the CNS in the alpha-synuclein mutant (A53T) mouse model. *J Neurosci* 2012; 2(49): 17775–17787.
69. Aguiar, AS, Duzzioni, M, and Remor, AP, et al. Moderate-intensity physical exercise protects against experimental 6-hydroxydopamine-induced hemiparkinsonism through Nrf2-antioxidant response element pathway. *Neurochem Res* 2015; 41(1–2): 64–72.
70. Boini, K.M., Singh, A. and Koka, S.S., 2021. Gut Microbial Metabolite Trimethylamine N-oxide Enhances Endoplasmic Reticular Stress and Promotes Endothelial Dysfunction. *Circulation*, 144(Suppl\_1), pp.A14071-A14071.
71. Wills, J, Jones, J, and Haggerty, T, et al. Elevated tauopathy and alpha-synuclein pathology in postmortem Parkinson's disease brains with and without dementia. *Exp Neurol* 2010; 225(1): 210–218.
72. Zhang, C, Li, C, and Chen, S, et al. Berberine protects against 6-OHDA-induced neurotoxicity in PC12 cells and zebrafish through hormetic mechanisms involving PI3K/AKT/Bcl-2 and Nrf2/HO-1 pathways. *Redox Biol* 2017; 11: 1–11.
73. Bresciani A, Missineo A, and Gallo, M, et al. Nuclear factor (erythroid-derived 2)-like 2 (NRF2) drug discovery: Biochemical toolbox to develop NRF2 activators by reversible binding of Kelch-like ECH-associated protein 1 (KEAP1). *Arch Biochem Biophys* 2017;631:31–41. doi: 10.1016/j.abb.2017.08.003
74. Wang L, Cai X, and Shi, M, et al. Identification and optimization of piperine analogues as neuroprotective agents for the treatment of Parkinson's disease via the activation of Nrf2/keap1 pathway. *Eur J Med Chem* 2020; 199: 112385. <https://doi.org/10.1016/j.ejmech.2020.112385>